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CD40.USPT.	390
CD40S	0
LIGAND.USPT.	29380
LIGANDS.USPT.	23963
GP39.USPT.	59
GP39S	0
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1 of 2

<u>L1</u>

42

Mar. 27, 1995 DATE FILED: APPL-NO: 08/411,098

L3: 5 of 10.

Eukaryotic layered vector initiation systems TITLE:

DATE ISSUED: Sep. 29, 1998 5,814,482 US PAT NO:

[ PMAGE AVAILABLE ]

Oct. 30, 1996 DATE FILED: 08/739,158 APPL-NO: Division of Ser. No. 404,796, Mar. 15, 1995, which is a REL-US-DATA:

continuation-in-part of Ser. No. 376,184, Jan. 18, 1995, abandoned, which is a continuation-in-part of Ser. No.

348,472, Nov. 30, 1994, abandoned, which is a

continuation-in-part of Ser. No. 198,450, Feb. 18, 1994, abandoned, which is a continuation-in-part of Ser. No.

122,791, Sep. 15, 1993, abandoned.

L3: 6 of 10

Fragments of a lymphocyte adhesion receptor for high TITLE:

endothelium, CD44

Sep. 15, 1998 DATE ISSUED: 5,808,004 US PAT NO:

[IMAGE AVAILABLE]

Jun. 7, 1995 DATE FILED: 08/472,543 APPL-NO:

Division of Ser. No. 884,624, May 15, 1992, Pat. No. REL-US-DATA: 5,504,194, which is a continuation of Ser. No. 628,646, Dec. 12, 1990, abandoned, which is a division of Ser.

No. 325,224, Mar. 17, 1989, Pat. No. 5,002,873.

L3: 7 of 10

Alphavirus structural protein expression cassettes TITLE: DATE ISSUED: Aug. 4, 1998 5,789,245

US PAT NO:

[IMAGE AVAILABLE] Oct. 30, 1996 DATE FILED: 08/741,881 APPL-NO:

Division of Ser. No. 404,796, Mar. 15, 1995, which is a REL-US-DATA: continuation-in-part of Ser. No. 376,184, Jan. 20, 1995,

abandoned, which is a continuation-in-part of Ser. No. 348,472, Nov. 30, 1994, abandoned, which is a

continuation-in-part of Ser. No. 198,450, Feb. 18, 1994, abandoned, which is a continuation-in-part of Ser. No.

122,791, Sep. 15, 1993, abandoned.

L3: 8 of 10

Diagnostic and therapeutic agents using a lymphocyte TITLE:

adhesion receptor for high endothelium CD44

Jun. 23, 1998 DATE ISSUED: 5,770,569 US PAT NO:

[IMAGE AVAILABLE]

Jun. 7, 1995 DATE FILED: 08/472,542 APPL-NO:

Division of Ser. No. 884,624, May 15, 1992, Pat. No. REL-US-DATA:

5,504,194, which is a continuation of Ser. No. 628,646, Dec. 12, 1990, abandoned, which is a division of Ser.

No. 325,224, Mar. 17, 1989, Pat. No. 5,002,873.

L3: 9 of 10

Lymphocyte adhesion receptor for high endothelium, CD44 TITLE:

DATE ISSUED: Apr. 2, 1996 5,504,194 US PAT NO: [IMAGE AVAILABLE]

May 15, 1992 DATE FILED: 07/884,624 APPL-NO:

Continuation of Ser. No. 628,646, Dec. 12, 1990,

REL-US-DATA: abandoned, which is a division of Ser. No. 325,224, Mar.

17, 1989, Pat. No. 5,002,873.

L3: 10 of 10

DNA sequence encoding a lymphocyte adhesion receptor for TITLE:

high endothelium

DATE ISSUED: Mar. 26, 1991 5,002,873 US PAT NO:

[IMAGE AVAILABLE]

Mar. 17, 1989 DATE FILED: 07/325,224 APPL-NO:

5,861,310 [IMAGE AVAILABLE] US PAT NO:

L3: 1 of 10

# ABSTRACT:

Tumor cells modified to express one or more T cell costimulatory molecules are disclosed. Preferred costimulatory molecules are B7-2 and B7-3. The tumor cells of the invention can be modified by transfection with nucleic acid encoding B7-2 and/or B7-3, by using an agent which induces or increases expression of B7-2 and/or B7-3 on the tumor cell or by coupling B7-2 and/or B7-3 to the tumor cell. Tumor cells modified to express B7-2 and/or B7-3 can be further modified to express B7. Tumor cells further modified to express MHC class I and/or class II molecules or in which expression of an MHC associated protein, the invariant chain, is inhibited are also disclosed. The modified tumor cells of the invention can be used in methods for treating a patient with a tumor, preventing or inhibiting metastatic spread of a tumor or preventing or inhibiting recurrence of a tumor. A method for specifically inducing a CD4.sup.+ T cell response against a tumor and a method for treating a tumor by modification of tumor cells in vivo are disclosed.

#### SUMMARY:

# BSUM(9)

Accordingly, the invention pertains to methods of inducing or enhancing T lymphocyte-mediated anti-tumor immunity in a subject by use of a modified tumor cell having increased immunogenicity. In one aspect of the invention, a tumor cell is modified to express one or more T cell costimulatory molecules on its surface. Preferred costimulatory molecules are novel B lymphocyte antigens, B7-2 and B7-3. Prior to modification, the tumor cell may lack the ability to express B7-2 and/or B7-3, may be capable of expressing B7-2 and/or B7-3 but fail to do so, or may express insufficient amounts of B7-2 and/or B7-3 to activate T cells. Therefore, a tumor cell can be modified by providing B7-2 and/or B7-3 to the tumor cell surface, by inducing the expression of B7-2 and/or B7-3 on the tumor cell or by increasing the level of expression of B7-2 and/or B7-3 on the tumor cell. In one embodiment, the tumor cell is modified by transfecting the cell with at least one nucleic acid encoding B7-2 and/or B7-3 in a form suitable for expression of the molecule(s) on the cell surface. Alternatively, the tumor cell is contacted with an agent which induces or increases expression of B7-2 and/or B7-3 on the cell surface. In yet another embodiment, the tumor cell is modified by chemically coupling B7-2 and/or B7-3 to the tumor cell surface. A tumor cell modified to express B7-2 and/or B7-3 can be further modified to express the T cell costimulatory molecule B7.

# SUMMARY:

# BSUM(10)

Even when provided with the ability to trigger a costimulatory signal in T cells, modified tumor cells may still be incapable of inducing anti-tumor T cell-mediated immune responses due to a failure to sufficiently trigger an antigen-specific primary activation signal. This can result from insufficient expression of MHC class I or class II molecules on the tumor cell surface. Accordingly, this invention encompasses modified tumor cells which provide both a T cell costimulatory signal and an antigen-specific primary activation signal, via an antigen-MHC complex, to T cells. Prior to modification, a tumor cell may lack the ability to express one or more MHC

' molecules, may be capable of expressing one or more MHC molecules but fail to do so, may express only certain types of MHC molecules (e.g., class I but not class II), or may express insufficient amounts of MHC molecules to activate T cells. Thus, in one embodiment, a tumor cell is modified by providing one or more MHC molecules to the tumor cell surface, by inducing the expression of one or more MHC molecules on the tumor cell surface or by increasing the level of expression of one or more MHC molecules on the tumor cell surface. Tumor cells expressing B7-2 and/or B7-3 are further modified, for example, by transfection with a nucleic acid encoding one or more MHC molecules in a form suitable for expression of the MHC molecule(s) on the tumor cell surface. Alternatively, such tumor cells are modified by contact with an agent which induces or increases expression of one or more MHC molecules on the cell.

DETDESC:

DETD(8)

The inability of a tumor cell to trigger a costimulatory signal in T cells may be due to a lack of expression of a costimulatory molecule, failure to express a costimulatory molecule even though the tumor cell is capable of expressing such a molecule, insufficient expression of a costimulatory molecule on the tumor cell surface or lack of expression of an appropriate costimulatory molecule (e.g. expression of B7 but not B7-2 and/or B7-3). Thus, according to one aspect of the invention, a tumor cell is modified to express B7-2 and/or B7-3 by transfection of the tumor cell with a nucleic acid encoding B7-2 and/or B7-3 in a form suitable for expression of B7-2 and/or B7-3 on the tumor cell surface. Alternatively, the tumor cell is modified by contact with an agent which induces or increases expression of B7-2 and/or B7-3 on the tumor cell surface. In yet another embodiment, B7-2 and/or B7-3 is coupled to the surface of the **tumor** cell to produce a modified tumor cell.

DETDESC:

A. Transfection of a Tumor Cell with a Nucleic Acid Encoding a Costimulatory Molecule

DETDESC:

DETD (15)

Alternatively, B7-2 and/or B7-3 can be expressed on a tumor cell using a plasmid expression vector which contains nucleic acid, e.g. a cDNA, encoding B7-2 and/or B7-3. Suitable plasmid expression. . . al., EMBO J. 6, 187-195 (1987)). Since only a small fraction of cells (about 1 out of 10.sup.5) typically integrate transfected plasmid DNA into their genomes, it is advantageous to transfect a nucleic acid encoding a selectable marker into the **tumor** cell along with the nucleic acid(s) of interest. Preferred selectable markers include those which confer resistance to drugs such as. . . on the same plasmid as the gene(s) of interest or may be introduced on a separate plasmid. Following selection of transfected tumor cells using the appropriate selectable marker(s), expression of the costimulatory molecule on the surface of the tumor cell can be confirmed by immunofluorescent staining of the cells. For example, cells may be stained with a fluorescently labeled monoclonal antibody reactive against the costimulatory molecule or with a fluorescently labeled soluble receptor which binds the costimulatory molecule. Expression of the B7-3 costimulatory molecule can be determined using a monoclonal antibody, BB1, which recognizes B7-3. Yokochi, T., et

al. J. Immunol. 128, 823-827 (1982).. . .

DETDESC:

DETD(19)

Another agent which can be used to induce or increase expression of B7-2 and/or B7-3 on a tumor cell surface is a nucleic acid encoding a transcription factor which upregulates transcription of the gene encoding the costimulatory molecule. This nucleic acid can be transfected into the tumor cell to cause increased transcription of the costimulatory molecule gene, resulting in increased cell-surface levels of the costimulatory molecule.

DETDESC:

DETD (24)

Before modification, a tumor cell may not express any costimulatory molecules, or may express certain costimulatory molecules but not others. As described herein, tumor cells can be modified by transfecting the tumor cell with nucleic acid encoding a costimulatory molecule(s), by inducing the expression of a costimulatory molecule(s) or by coupling a costimulatory molecule(s) to the tumor cell. For example, a tumor cell transfected with nucleic acid encoding B7-2 can be further transfected with nucleic acid encoding B7. The cDNA sequence and deduced amino acid sequence of human and mouse B7 is shown. . . and SEQ ID NO:7 and 8, respectively. Alternatively, more than one type of modification can be used. For example, a tumor cell transfected with a nucleic acid encoding B7-2 can be stimulated with an agent which induces expression of B7.

DETDESC:

DETD (26)

Another aspect of this invention features modified tumor cells which express a costimulatory molecule and which express one or more MHC molecules on their surface to trigger both a costimulatory signal and a primary, antigen-specific, signal in T cells. Before modification, tumor cells may be unable to express MHC molecules, may fail to express MHC molecules although they are capable of expressing such molecules, or may express insufficient amounts of MHC molecules on the tumor cell surface to cause T cell activation. Tumor cells can be modified to express either MHC class I or MHC class II molecules, or both. One approach to modifying tumor cells to express MHC molecules is to transfect the tumor cell with one or more nucleic acids encoding one or more MHC molecules. Alternatively, an agent which induces or increases expression of one or more MHC molecules on tumor cells can be used to modify tumor cells. Inducing or increasing expression of MHC class II molecules on a tumor cell can be particularly beneficial for activating CD4.sup.+ T cells against the tumor since the ability of MHC class II.sup.+ tumor cells to directly present tumor peptides to CD4.sup.+ T cells bypasses the need for professional MHC class II.sup.+ APCs. This can improve tumor immunogenicity because soluble tumor antigen (in the form of tumor cell debris or secreted protein) may not be available for uptake by professional MHC class II.sup.+ APCs.

DETDESC:

DETD(31)

Fragments, mutants or variants of MHC class II molecules that retain

the ability to bind peptide antigens and activate T cell responses, as evidenced by proliferation and/or lymphokine production by T cells, are considered within the scope of the invention. A preferred variant is an MHC class II molecule in which the cytoplasmic domain of either one or both of the .alpha. and .beta. chains is truncated. It is known that truncation of the cytoplasmic domains allows peptide binding by and  $\operatorname{cell}$ surface expression of MHC class II molecules but prevents the induction of endogenous B7 expression, which is triggered by an intracellular signal generated by the cytoplasmic domains of the MHC class II protein chains upon crosslinking of cell surface MHC class II molecules. Kuolova. L., et al., J. Exp. Med. 173, 759-762 (1991); Nabavi, N., et al. Nature 360, 266-268 (1992). Expression of B7-2 and B7-3 is also induced by crosslinking surface MHC class II molecules, and thus truncation of MHC class II molecules may also prevent induction of B7-2 and/or B7-3. In tumor cells transfected to constitutively express B7-2 and/or B7-3, it may be desirable to inhibit the expression of endogenous costimulatory molecules, for instance to restrain potential downregulatory feedback mechanisms. Transfection of a tumor cell with a nucleic acid(s) encoding a cytoplasmic domain-truncated form of MHC class II .alpha. and .beta. chain proteins would.

## DETDESC:

## DETD (56)

The modified tumor cells of the present invention can be used to increase tumor immunogenicity, and therefore can be used therapeutically for inducing or enhancing T lymphocyte-mediated anti-tumor immunity in a subject with a tumor or at risk of developing a tumor. A method for treating a subject with a tumor involves obtaining tumor cells from the subject, modifying the tumor cells ex vivo to express a T cell costimulatory molecule, for example by transfecting them with an appropriate nucleic acid, and administering a therapeutically effective dose of the modified tumor cells to the subject. Appropriate nucleic acids to be introduced into a tumor cell include nucleic acids encoding B7-2 and/or B7-3, alone or together with nucleic acids encoding B7, MHC molecules (class I or class II) or Ii antisense sequences as described herein. Alternatively, after tumor cells are obtained from a subject, they can be modified ex vivo using an agent which induces or increases expression of B7-2 and/or B7-3 (and possibly also using agent(s) which induce or increase B7 or MHC molecules).

US PAT NO: 5,858,776 [IMAGE AVAILABLE] L3: 2 of 10

#### ABSTRACT:

Tumor cells modified to express a T cell costimulatory molecule are disclosed. In one embodiment, the costimulatory molecule is a CD28/CTLA4 ligand, preferably a B lymphocyte antigen B7. The tumor cells of the invention can be modified by transfection with nucleic acid encoding a T cell costimulatory molecule, by using an agent which induces or increases expression of a T cell costimulatory molecule on the tumor cell surface or by coupling a T cell costimulatory molecule to the tumor cell surface. Tumor cells further modified to express MHC class I and/or class II molecules or in which expression of an MHC associated protein, the invariant chain, is inhibited are also disclosed. The modified tumor cells of the invention can be used in methods for treating a patient with a tumor, preventing or inhibiting metastatic spread of a tumor or preventing or inhibiting recurrence of a tumor. A method for specifically inducing a CD4.sup.+ T cell response against a tumor and a method for treating a tumor by modification of tumor cells in vivo are disclosed.

SUMMARY:

BSUM(9)

Accordingly, the invention pertains to methods of inducing or enhancing T lymphocyte-mediated anti-tumor immunity in a subject by use of a modified tumor cell having increased immunogenicity. In one aspect of the invention, a tumor cell is modified to express a T cell costimulatory molecule on its surface. Prior to modification, the tumor cell may lack the ability to express a T cell costimulatory molecule, may be capable of expressing a T cell costimulatory molecule but fail to do so, or may express insufficient amounts of a T cell costimulatory molecule to activate T cells. Therefore, a tumor cell can be modified by providing a costimulatory molecule to the tumor cell surface, by inducing the expression of a costimulatory molecule on the tumor cell surface or by increasing the level of expression of a costimulatory molecule on the tumor cell surface. In one embodiment, the tumor cell is modified by transfecting the cell with a nucleic acid encoding a T cell costimulatory molecule in a form suitable for expression of the molecule on the cell surface. Alternatively, the tumor cell is contacted with an agent which induces or increases expression of a T cell costimulatory molecule on the cell surface. In yet another embodiment, the tumor cell is modified by chemically coupling a T cell costimulatory molecule to the tumor cell surface.

SUMMARY:

BSUM(11)

Even when provided with the ability to trigger a costimulatory signal in T cells, modified tumor cells may still be incapable of inducing anti-tumor T cell-mediated immune responses due to a failure to sufficiently trigger an antigen-specific primary activation signal. This can result from insufficient expression of MHC class I or class II molecules on the tumor cell surface. Accordingly, this invention encompasses modified tumor cells which provide both a T cell costimulatory signal and an antigen-specific primary activation signal, via an antigen-MHC complex, to T cells. Prior to modification, a . tumor cell may lack the ability to express one or more MHC molecules, may be capable of expressing one or more MHC molecules but fail to do so, may express only certain types of MHC molecules (e.g., class I but not class II), or may express insufficient amounts of MHC molecules to activate T cells. Thus, in one embodiment, a tumor cell is modified by providing one or more MHC molecules to the tumor cell surface, by inducing the expression of one or more MHC molecules on the tumor cell surface or by increasing the level of expression of one or more MHC molecules on the tumor cell surface. Tumor cells expressing a T cell costimulatory molecule are further modified, for example, by transfection with a nucleic acid encoding one or more MHC molecules in a form suitable for expression of the MHC molecule(s) on the tumor cell surface. Alternatively, such tumor cells are modified by contact with an agent which induces or increases expression of one or more MHC molecules on the cell.

DETDESC:

DETD(5)

The inability of a tumor cell to trigger a costimulatory signal in T cells may be due to a lack of expression of a costimulatory molecule, failure to express a costimulatory molecule even though the tumor cell is capable of expressing such a molecule, or insufficient expression of a costimulatory molecule on the

tumor cell surface. Thus, according to one aspect of the invention, a tumor cell is modified to express a costimulatory molecule by transfection of the tumor cell with a nucleic acid encoding a costimulatory molecule in a form suitable for expression of the costimulatory molecule on the tumor cell surface. Alternatively, the tumor cell is modified by contact with an agent which induces or increases expression of a costimulatory molecule on the tumor cell surface. In yet another embodiment, a costimulatory molecule is coupled to the surface of the tumor cell to produce a modified tumor cell. The term "costimulatory molecule" is defined herein as a molecule which interacts with a T cell which has received a primary activation signal to result in T cell proliferation and/or cytokine production. Preferred costimulatory molecules include antigens on the surface of B lymphocytes, professional antigen presenting cells (e.g., monocytes, dendritic cells, Langerhans cells) and other. . . CD28, CTLA4, both CD28 and CTLA4, or other known or as yet undefined receptors on immune cells. A particularly preferred costimulatory molecule which binds CD28 and/or CTLA4 is the B lymphocyte antigen B7.

#### DETDESC:

# DETD(8)

A. Transfection of a Tumor Cell with a Nucleic Acid Encoding a Costimulatory Molecule

DETDESC:

DETD(9)

Tumor cells can be modified ex vivo to express a T cell costimulatory molecule by transfection of isolated tumor cells with a nucleic acid encoding a costimulatory molecule in a form suitable for expression of the molecule on the surface of the tumor cell. The terms "transfection" or "transfected with" refers to the introduction of exogenous nucleic acid into a mammalian cell and encompass a variety of techniques useful. . . acids into mammalian cells including electroporation, calcium-phosphate precipitation, DEAE-dextran treatment, lipofection, microinjection and infection with viral vectors. Suitable methods for transfecting mammalian cells can be found in Sambrook et al. (Molecular Cloning: A Laboratory Manual, 2nd Edition, Cold Spring Harbor Laboratory. . . and other laboratory textbooks. The nucleic acid to be introduced can be, for example, DNA encompassing the gene encoding the costimulatory molecule, sense strand RNA encoding the costimulatory molecule or a recombinant expression vector containing a cDNA encoding the costimulatory molecule. Preferred cDNAs to use are those for human and mouse B7 (Freeman, G. J., et al., J Exp. Med 174,. .

## DETDESC:

#### DETD(12)

Alternatively, a costimulatory molecule can be expressed on a tumor cell using a plasmid expression vector which contains nucleic acid, e.g. a cDNA, encoding the costimulatory molecule. Suitable plasmid expression vectors include CDM8 (Seed, B., Nature 329, 840(1987)) and pMT2PC (Kaufman, et al., EMBO J 6, 187-195 (1987)). Since only a small fraction of cells (about 1 out of 10.sup.5) typically integrate transfected plasmid DNA into their genomes, it is advantageous to transfect a nucleic acid encoding a selectable marker into the tumor cell along with the nucleic acid(s) of interest. Preferred selectable markers include those which confer resistance to drugs such as. . . on the same plasmid as the gene(s) of interest or may be

introduced on a separate plasmid. Following selection of transfected tumor cells using the appropriate selectable marker(s), expression of the costimulatory molecule on the surface of the tumor cell can be confirmed by immunofluorescent staining of the cells. For example, cells may be stained with a fluorescently labeled monoclonal antibody reactive against the costimulatory molecule or with a fluorescently labeled soluble receptor which binds the costimulatory molecule. Expression of the B7 costimulatory molecule can be determined using a monoclonal antibody, 133, which recognizes B7. Freedman, A. S., et al. J Immunol. 139, 3260-3267. . .

## DETDESC:

#### DETD(13)

When transfection of tumor cells leads to modification of a large proportion of the tumor cells and efficient expression of a costimulatory molecule on the surface of tumor cells, e.g. when using a viral expression vector, tumor cells may be used without further isolation or subcloning. Alternatively, a homogenous population of transfected tumor cells can be prepared by isolating a single transfected tumor cell by limiting dilution cloning followed by expansion of the single tumor cell into a clonal population of cells by standard techniques.

#### DETDESC:

## DETD(16)

Another agent which can be used to induce or increase expression of a costimulatory molecule on a tumor cell surface is a nucleic acid encoding a transcription factor which upregulates transcription of the gene encoding the costimulatory molecule. This nucleic acid can be transfected into the tumor cell to cause increased transcription of the costimulatory molecule gene, resulting in increased cell-surface levels of the costimulatory molecule.

# DETDESC:

#### DETD (20)

Another aspect of this invention features modified tumor cells which express a costimulatory molecule and which express one or more MHC molecules on their surface to trigger both a costimulatory signal and a primary, antigen-specific, signal in T cells. Before modification, tumor cells may be unable to express MHC molecules, may fail to express MHC molecules although they are capable of expressing such molecules, or may express insufficient amounts of MHC molecules on the tumor cell surface to cause T cell activation. Tumor cells can be modified to express either MHC class I or MHC class II molecules, or both. One approach to modifying tumor cells to express MHC molecules is to transfect the tumor cell with one or more nucleic acids encoding one or more MHC molecules. Alternatively, an agent which induces or increases expression of one or more MHC molecules on tumor cells can be used to modify tumor cells. Inducing or increasing expression of MHC class II molecules on a tumor cell can be particularly beneficial for activating CD4.sup.+ T cells against the tumor since the ability of MHC class II.sup.+ tumor cells to directly present tumor peptides to CD4.sup.+ T cells bypasses the need for professional MHC class II.sup.+ APCs. This can improve tumor immunogenicity because soluble tumor antigen (in the form of tumor cell debris or secreted protein) may not be available for uptake by professional MHC class II.sup.+ APCs.

#### DETDESC:

Fragments, mutants or variants of MHC class II molecules that retain the ability to bind peptide antigens and activate T cell responses, as evidenced by proliferation and/or lymphokine production by T cells, are considered within the scope of the invention. A preferred variant is an MHC class II molecule in which the cytoplasmic domain of either one or both of the .alpha. and .beta. chains is truncated. Truncation of the cytoplasmic domains allows peptide binding by and cell surface expression of MHC class II molecules but prevents the induction of endogenous B7 expression, which is triggered by an intracellular signal generated by the cytoplasmic domains of the MHC class II protein chains upon crosslinking of cell surface MHC class II molecules. Kuolova. L., al., J. Exp. Med. 173, 759-762 (1991); Nabavi, N., et al. Nature 360, 266-268 (1992). In tumor cells transfected to constitutively express B7 or other costimulatory molecule, it may be desirable to inhibit the expression of endogenous B7, for instance to restrain potential downregulatory feedback mechanisms. Transfection of a tumor cell with a nucleic acid(s) encoding a cytoplasmic domain-truncated form of MHC class II .alpha. and .beta. chain proteins would.

## DETDESC:

#### DETD (35)

The tumor cells to be modified as described herein include tumor cells which can be transfected or treated by one or more of the approaches encompassed by the present invention to express a costimulatory molecule. If necessary, the tumor cell can be further modified to express MHC molecules or an inhibitor of Ii expression. A tumor from which tumor cells are obtained can be one that has arisen spontaneously, e.g in a human subject, or may be experimentally derived or induced, e.g. in an animal subject. The tumor cells can be obtained, for example, from a solid tumor of an organ, such as a tumor of the lung, liver, breast, colon, bone etc. Malignancies of solid organs include carcinomas, sarcomas, melanomas and neuroblastomas. The tumor cells can also be obtained from a blood-borne (ie. dispersed) malignancy such as a lymphoma, a myeloma or a leukemia.

#### DETDESC:

#### DETD (36)

The tumor cells to be modified include those that express MHC molecules on their cell surface prior to transfection and those that express no or low levels of MHC class I and/or class II molecules. A minority of normal cell types express MHC class II molecules. It is therefore expected that many tumor cells will not express MHC class II molecules naturally. These tumors can be modified to express a costimulatory molecule and MHC class II molecules. Several types of tumors have been found to naturally express surface MHC class II molecules, such as melanomas (van Duinen et al., Cancer Res. 48, 1019-1025, 1988), diffuse large cell lymphomas (O'Keane et al., Cancer 66, 1147-1153, 1990), squamous cell carcinomas of the head and neck (Mattijssen et al., Int. J Cancer 6, 95-100, 1991) and colorectal carcinomas (Moller et al., Int. J Cancer 6, 155-162, 1991). Tumor cells which naturally express class II molecules can be modified to express a costimulatory molecule, and, in addition, other class II molecules which can increase the spectrum of TAA peptides which can be presented by the tumor cell. Most non-malignant cell types express MHC class I molecules. However, malignant transformation is often accompanied by

downregulation of expression of MHC class I molecules on the surface of tumor cells. Csiba, A., et al., Brit. J Cancer 50, 699-709 (1984). Importantly, loss of expression of MHC class I antigens by tumor cells is associated with a greater aggressiveness and/or metastatic potential of the tumor cells. Schrier, P. I., et al. Nature 305, 771-775 (1983); Holden, C. A., et al. J Am. Acad. Dermatol. 9., 867-871 (1983); Baniyash, M., et al. J Immunol. 129, 1318-1323 (1982). Types of tumors in which MHC class I expression has been shown to be inhibited include melanomas, colorectal carcinomas and squamous cell carcinomas. van Duinen et al., Cancer Res. 48, 1019-1025, (1988); Moller et al., Int. J Cancer 6, 155-162, (1991); Csiba, A., et al., Brit. J Cancer 50, 699-709 (1984); Holden, C. A., et al. J Am. Acad. Dermatol. 9., 867-871 (1983). A tumor cell which fails to express class I molecules or which expresses only low levels of MHC class I molecules can be modified by one or more of the techniques described herein to induce or increase expression of MHC class I molecules on the tumor cell surface to enhance tumor cell immunogenicity.

## DETDESC:

DETD (49)

The modified tumor cells of the present invention can be used to increase tumor immunogenicity, and therefore can be used therapeutically for inducing or enhancing T lymphocyte-mediated anti-tumor immunity in a subject with a tumor or at risk of developing a tumor. A method for treating a subject with a tumor involves obtaining tumor cells from the subject, modifying the tumor cells ex vivo to express a T cell costimulatory molecule, for example by transfecting them with an appropriate nucleic acid, and administering a therapeutically effective dose of the modified tumor cells to the subject. Appropriate nucleic acids to be introduced into a tumor cell include a nucleic acid encoding a T cell costimulatory molecule, for example a CD28 and/or CTLA4 ligand such as B7, alone or together with nucleic acids encoding MHC molecules (class I or class II) or Ii antisense sequences as described herein. Alternatively, after tumor cells are obtained from a subject, they can be modified ex vivo using an agent which induces or increases expression of a costimulatory molecule (and possibly also using agent(s) which induce or increase MHC molecules).

US PAT NO: 5,843,723 [IMAGE AVAILABLE] L3: 3 of 10

DETDESC:

DETD (72)

Another example of an immunomodulatory cofactor is the B7/BB1 costimulatory factor. Briefly, activation of the full functional activity of  $ar{ t T}$  cells requires two signals. One signal is provided by interaction of the antigen-specific T cell receptor with peptides which are bound to major histocompatibility complex (MHC) molecules, and the second signal, referred to as costimulation, is delivered to the T cell by antigen-presenting cells. Briefly, the second signal is required for interleukin-2 (IL-2) production by T cells and appears to involve interaction of the B7/BB1 molecule on antigen-presenting cells with CD28 and CTLA-4 receptors on T lymphocytes (Linsley et al., J. Exp. Med., 173:721-730, 1991a, and J. Exp. Med., 174:561-570, 1991). Within one embodiment of the invention, B7/BB1 may be introduced into tumor cells in order to cause costimulation of CD8.sup.+ T cells, such that the CD8.sup.+ T cells produce enough IL-2 to expand and become fully activated. These CD8.sup.+ T cells can kill tumor cells that are not expressing B7 because costimulation is no longer required for further CTL function. Vectors that express both the costimulatory B7/BB1 factor

and, for example, an immunogenic HBV core protein, may be made utilizing methods which are described herein. Cells transduced with these vectors will become more effective antigen-presenting cells. The HBV core-specific CTL response will be augmented from the fully activated CD8.sup.+ T cell via the costimulatory ligand B7/BB1.

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L1
              6 S E4, E5
             61 S (ACCESSORY OR COSTIMULATORY) (P) (MOLECULE?) (P) (TRANSDUC?
L2
OR
             10 S L2(P) (TUMOR? OR TUMOUR? OR CANCER)
L3
=> s 12 and (cd40(w)ligand or cd40L or gp39)
           165 CD40
         21239 LIGAND
            71 CD40(W)LIGAND
            20 CD40L
            22 GP39
             0 L2 AND (CD40(W)LIGAND OR CD40L OR GP39)
L4
=> s (tranfect? or transduc?)(P)(cd40(w)ligand or cd40L or gp39)
           159 TRANFECT?
         97056 TRANSDUC?
           165 CD40
         21239 LIGAND
            20 CD40L
             3 (TRANFECT? OR TRANSDUC?) (P) (CD40(W) LIGAND OR CD40L OR GP39)
T.5
=> d 15 1-3 date
                                                         L5: 1 of 3
TITLE:
              TRAF inhibitors
                                         DATE ISSUED:
                                                        Aug. 4, 1998
               5,789,550
US PAT NO:
               [IMAGE AVAILABLE]
                                                        Aug. 14, 1996
                                         DATE FILED:
APPL-NO:
               08/700,749
                                                         L5: 2 of 3
               Product and process for targeting an immune response
TITLE:
                                         DATE ISSUED:
                                                        Dec. 16, 1997
US PAT NO:
               5,698,679
               [IMAGE AVAILABLE]
                                                         Sep. 19, 1994
                                         DATE FILED:
               08/309,006
APPL-NO:
                                                         L5: 3 of 3
               Method of preventing or treating disease characterized by
TITLE:
                 neoplastic cells expressing CD40
               5,674,492
                                         DATE ISSUED:
                                                         Oct. 7, 1997
US PAT NO:
               [IMAGE AVAILABLE]
                                         DATE FILED:
                                                        Dec. 21, 1994
APPL-NO:
               08/360,923
               Continuation-in-part of Ser. No. 172,664, Dec. 23, 1993,
REL-US-DATA:
                 abandoned.
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L5: 1 of 3

=> d 15 1-3 kwic

US PAT NO: 5,789,550 [IMAGE AVAILABLE]

DETDESC:

DETD(84)

Compounds . . . agent, would release TRAF from its complexed form. As an example, this compound can be a native ligand the signal transduction of which is mediated by TRAF, e.g. TNF, a CD40 ligand, a CD30 ligand, etc. The concentration of the free or bound TRAF can then be detected and/or the dissociation constant.

L5: 2 of 3 5,698,679 [IMAGE AVAILABLE] US PAT NO:

DETDESC:

DETD(29)

. cause a T cell to be stimulated, anergized or killed. Activation of a T cell refers to induction of signal transduction pathways in the T cell resulting in production of cellular products (e.g., interleukin-2) by that T cell. Anergy refers to. . . antigen. Effector molecules involved in T cell activation include, but are not limited to, B7, B7-2, CD28, CD40 and the CD40 ligand. APCs having B7 or B7-2, and CD40 are capable of activating T cells by binding to CD28 and CD40 ligand, respectively, on the surface of a T cell. Such APCs are referred to as professional APCs. APCs lacking B7 or.

L5: 3 of 3 5,674,492 [IMAGE AVAILABLE] US PAT NO:

DETDESC:

DETD (23)

The . . . disclosed herein is a ligand for CD40, a receptor that is a member of the TNF receptor super family. Therefore, CD40L is likely to be responsible for transducing signal via CD40, which is known to be expressed, for example, by B lymphocytes. Full-length CD40L is a membrane-bound polypeptide with an extracellular region at its C terminus, a transmembrane region, and an intracellular region at its N-terminus. A soluble version of CD40L can be made from the extracellular region or a fragment thereof and a soluble CD40L has been found in culture supernatants from cells that express a membrane-bound version of CD40L. The protein sequence of the extracellular region of murine CD40L extends from amino acid 47 to amino acid 260 in SEQ ID NO:2 of USSN 07/969,703. The protein sequence of the extracellular region of human CD40L extends from amino acid 47 to amino acid 261 in SEQ ID NO:2. The biological activity of CD40L is mediated by binding to CD40or a species-specific homolog thereof and comprises proliferation of B cells and induction of immunoglobulin secretion from activated B cells. CD40L (including soluble monomeric and oligomeric forms, as well as membrane-bound forms) can effect B cell proliferation and immunoglobulin secretion (except.

=> s (tumor? or tumour? or cancer)(P)(cd40(w)ligand or cd40L or gp39)

26041 TUMOR? 2429 TUMOUR?

26098 CANCER

165 CD40

21239 LIGAND

20 CD40L

37 (TUMOR? OR TUMOUR? OR CANCER) (P) (CD40(W) LIGAND OR CD40L OR 22 GP39

L6 GP3

The  $\cdot$   $\cdot$  or negative apoptotic signal. For example, physiological stimuli that prevent or inhibit apoptosis include, for example, growth factors, extracellular matrix, CD40 ligand, viral gene products neutral amino acids, zinc, estrogen and androgens: In contrast, stimuli which promote apoptosis include growth factors such as tumor necrosis factor (TNF), Fas, and transforming growth factor .beta. (TGF.beta.), neurotransmitters, growth factor withdrawal, loss of extracellular matrix attachment, intracellular calcium and glucocorticoids, for example. Other.

US PAT NO:

5,837,816 [IMAGE AVAILABLE]

L7: 2 of 5

DETDESC:

DETD(2)

The present invention relates to a method of preparing a soluble hetero-oligomeric mammalian polypeptide (or protein) by culturing a host cell transformed or transfected with an expression vector encoding a fusion protein comprising a leucine zipper domain and a heterologous mammalian protein. Preferably, the. . . that act together to bind a ligand (such as IL-2), for example. Exemplary mammalian transmembrane proteins include members of the tumor necrosis factor/nerve growth factor receptor (TNFR/NGFR) family (Farrah and Smith, Nature 358:26, 1992; Goodwin et al., Cell 73:447; 1993), which includes CD40 Ligand (CD40-L), CD27 Ligand (CD27-L), OX40 Ligand (OX40-L), and TNF. Structural studies of certain members of this family of proteins indicate. .

US PAT NO:

5,786,173 [IMAGE AVAILABLE]

L7: 3 of 5

SUMMARY:

BSUM(5)

The . . or negative apoptotic signal. For example, physiological stimuli that prevent or inhibit apoptosis include, for example, growth factors, extracellular matrix, CD40 ligand, viral gene products neutral amino acids, zinc, estrogen and androgens. In contrast, stimuli which promote apoptosis include growth factors such as tumor necrosis factor (TNF), Fas, and transforming growth factor .beta. (TGF.beta.), neurotransmitters, growth factor withdrawal, loss of extracellular matrix attachment, intracellular calcium and glucocorticoids, for example.

US PAT NO:

5,716,805 [IMAGE AVAILABLE]

L7: 4 of 5

DETDESC:

DETD(2)

The present invention relates to a method of preparing a soluble mammalian protein by culturing a host cell transformed or transfected with an expression vector encoding a fusion protein comprising a zipper domain and a heterologous mammalian protein. In one embodiment,. . . heterologous mammalian protein comprises an extracellular domain of a mammalian transmembrane protein. Exemplary mammalian transmembrane proteins include members of the tumor necrosis factor/nerve growth factor receptor (TNFR/NGFR) family (Farrah and Smith, Nature 358:26, 1992; Goodwin et al., Cell 73:447; 1993), which includes CD40 Ligand (CD40-L), CD27 Ligand (CD27-L), OX40 Ligand (OX40-L), and TNF. Structural studies of certain members of this family of proteins indicate. .

US PAT NO:

5,674,492 [IMAGE AVAILABLE] L7: 5 of 5

DETDESC:

DETD (92)

This example illustrates the effect of recombinant human CD40 ligand on the growth of human B-cell lymphomas in SCID mice. SCID mice were obtained, and treated substantially as described in Example 4, above. On day 0, SCID mice were injected either intraperitoneally with 5.times.10.sup.6 RL or TU2C cells. The tumor cell recipients then received 100 .mu.l of concentrated supernatant fluid from cells transfected with either a vector encoding human CD40 ligand, or vector alone (control). Two concentrations of the CD40 ligand containing supernatant fluid were tested: a ten-fold concentrate and a two-fold concentrate (10x and 2x, respectively). The concentrated supernatants were administered. . . third day for a period of 15 days (total of 5 injections), starting at day 3. Mice were monitored for tumor development and progression; moribund mice were euthanized. All mice were necropsied for evidence of tumor. Liver, kidney and lymphoid organs were analyzed histologically for presence of tumor cells. Both parametric (student's t test) and non-parametric (Wilcoxan rank sum test) analyses were performed to determine if the groups. . .